USB® PrepEase® Gel Extraction Kit
Product numbers 78756/78757

Brief protocol for gel extraction

**Important:** Check that ethanol was added to NT3 Buffer before starting.

1. **Excise DNA fragment**
   a. Excise DNA from the agarose gel using TAE Buffer. Minimize the gel volume.
   b. Determine the weight of the gel slice.

2. **Gel solubilization**
   a. Add 200 µl NT Buffer for each 100 mg of agarose gel slice.
   b. Incubate sample at 50°C until gel is completely dissolved (5-10 minutes).
   c. Vortex sample briefly every 2-3 minutes.

3. **Bind DNA sample to column**
   a. Place PrepEase® Cleanup Column into a 2 ml PrepEase Collecting Tube.
   b. Load the sample directly into the center of the cleanup column.
   c. Centrifuge for 1 minute at 11,000 x g.
   d. Discard flow-through.

4. **Wash column**
   a. Add 600 µl NT3 Buffer to column.
   b. Centrifuge for 1 minute at 11,000 x g.
   c. Discard flow-through. Place cleanup column back into collecting tube.

5. **Dry column**
   a. Centrifuge for 2 minutes at 11,000 x g to remove excess NT3 Buffer.

6. **Elute DNA**
   a. Place the cleanup column into a clean 1.5 ml microcentrifuge tube.
   b. Add 15-50 µl NE Buffer to column.
   c. Incubate at room temperature for 1 minute.
   d. Centrifuge for 1 minute at 11,000 x g.

Brief protocol for PCR purification

**Important:** Check that ethanol was added to NT3 Buffer before starting.

1. **Prepare sample for binding**
   a. Add 2 volumes of NT Buffer per 1 volume of DNA containing sample. For example, add 200 µl NT Buffer to 100 µl of PCR mixture.

Continue with Step 3 of the protocol for DNA extraction from agarose gels.

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